Quick Guide: i-Ome® Protein Array Assay Procedure

Thaw and Dilute Samples

- Thaw samples at room temperature (RT): 20 – 22°C.
- Vortex the samples and centrifuge at 13,000 g for 3 minutes to pellet any debris.
- Dilute the samples by adding 22.5µL of sample into 4 mL of Serum Assay Buffer (1:200 dilution).

Sample Application

- Wash slides in chilled Serum Assay Buffer (SAB).
- Add samples to the CELLSTAR® FourWell plates. 4 mL of diluted sample per each well.
- Place the slides inside the FourWell plates and cover.

Sample Incubation

- Incubate the slides on an orbital shaker set to 50 rpm, for 1 hour at RT.

Washing after Sample Incubation

- Briefly wash slides (10 – 15 seconds) with SAB (200 mL) in a slide staining dish.
- Wash 3 times (5 minutes each wash) on an orbital shaker set to 50 rpm, at RT, in 200 mL of SAB in slide staining dish.

Cy3-Anti-Human IgG Incubation

- Dilute 1:1,000 Cy3-anti human-IgG (200 µL into 200 mL of SAB) and gently mix.
- Pour diluted Cy3-anti human-IgG into a clean slide staining dish.
- Place the slide rack into the dish containing 200 mL of diluted Cy3-anti-hIgG.
- Incubate on orbital shaker at 50 rpm at RT for 1 hour.

Wash after Cy3-Anti-Human IgG Incubation

- Wash the slides 3 times for 5 minutes on an orbital shaker in 200 mL of SAB in a slide staining dish.
- Rinse the slides 3 times with 200 mL of high purity water in a fresh dish.

Drying and Scanning Slides

- Air dry the slides overnight protected from light or by centrifugation at 400 g for 4 minutes.
- Scan slides using a microarray scanner, 16-bit, 10 µm resolution.